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### Review

## Neurogenesis in the epileptic brain: a brief overview from temporal lobe epilepsy

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#### Abstract:

Dentate granule cell neurogenesis persists throughout life in the hippocampus of mammals. Alterations in this process occur in many neurological diseases, including epilepsy. Among the different types of epilepsy, the most frequent is temporal lobe epilepsy (TLE). Therefore, a number of laboratory studies use animal models of TLE to observe the fate of neuronal cells after seizures. Hippocampal neurogenesis is very sensitive to physiological and pathological stimuli. Seizures, as pathological stimuli, alter both the extent and the pattern of neurogenesis, which is associated with cognitive function. Various alterations in neurogenesis are observed depending on the amount of time that has elapsed after the seizures. In acute seizures, neurogenesis generally increases, whereas in chronic epilepsy, neurogenesis decreases. Moreover, several methods currently used for the treatment of brain disorders such as TLE can also have significant impacts on cognitive functions. This review is focused on the recent findings regarding neurogenesis in animal models of TLE.

#### Key words:

dentate gyrus, seizures, pilocarpine, kainic acid, antiepileptic drugs

Abbreviations: AED(s) – antiepileptic drug(s), DG – dentate gyrus, KA – kainic acid, MGE – medial ganglionic eminence, PILO – pilocarpine, SE – status epilepticus, SGZ – subgranular zone, SRMS – spontaneous recurrent motor seizures, SVZ – subventricular zone, TLE – temporal lobe epilepsy

## Introduction

Neurogenesis is a developmental process that involves the proliferation, migration and differentiation of neuroblasts and the synaptic integrations of newborn neurons. Proliferation of new cells continues into old age, even in humans [20]. There are two major neurogenic regions in the brain: the dentate gyrus (DG) of the hippocampus and the subventricular zone (SVZ) of the brain [38, 44]. The neurogenic cells of the SVZ may constitute a population of undifferentiated cells that can be recruited after tissue injury [19, 45, 48]. The newly born granule cells in the DG have been extensively studied over the last several years. They send axonal projections to their normal target zone, the mossy fiber pathway [27]. Their dendritic trees resemble those of other granule cells, although some aspects of their dendrites and spines have been suggested to be immature [51]. Moreover, the newly born granule cells appear to develop electrophysiological properties similarly to other granule cells [77]. The functional integration of newly born dentate granule cells into the hippocampal circuitry [35, 60] and their ability to mediate long-term potentiation in DG [63] has led to the hypothesis that neurogenesis in the adult brain may play a key role in learning and memory [67, 69], as well as cognitive dysfunction in some diseases, such as temporal lobe epilepsy (TLE), Alzheimer's disease and major depressive disorders. Hippocampal neurogenesis is very sensitive to physiological and pathological stimuli. Seizures, as pathological stimuli, alter both the extent and the pattern of neurogenesis, although the overall effects depend on the type of seizure [40].

## Animal models of TLE

Epilepsy is one of the oldest and most well-known ailments of the brain, affecting 50 million people worldwide [58]. According to epidemiological studies, approximately 70–80% of epilepsy patients achieve remission, but there are still patients who are refractory to currently available treatments [41, 46, 57].

Understanding the molecular mechanisms associated with seizure development can be addressed by dividing all in vivo animal models into two categories: models of seizures and models of epilepsy. The difference between these two groups is those models of epilepsy are characterized by multiple spontaneous recurrent seizures (TLE evoked by pilocarpine or kainic acid), whereas models of seizures are characterized by generalized seizures in response to a single exposure to a potent neurotoxin [3, 47]. The inherent distinction between the two types of models might foster a better understanding of critical elements in the evolution of seizures. Among the many different animal models of epilepsy, the most well known and most frequently used is TLE. Because human TLE is the most common type of epilepsy, animal models of this condition are thought to be some of the best for helping us understand the problem of epileptogenesis and the neuronal alterations that take place in the brain after convulsions. Several well-characterized animal models of TLE exist, and they reflect, at least in part, the complex partial seizures observed in patients with TLE [59]. Administration of chemical convulsants, including the glutamate analogue kainic acid (KA) and the cholinergic agonist pilocarpine (PILO), and electrical stimulation of the amygdala or hippocampus (kindling model) are the most frequently studied animal models of TLE. PILO and KA can be systemically or intracerebrally injected into animals and can rapidly produce seizures with an acute episode of status epilepticus (SE). The majority of rodents that survive the initial SE develop spontaneous seizures after a quiescent period of several days to several weeks [11]. The PILO model is well characterized in rats [12, 74] and in various strains of mice [68, 73]. According to the results from many different studies, PILO induces neurochemical alterations in neurons and glial cells, which in turn change the cellular environment by altering the expression levels of receptors, trophic factors, enzymes and proteins from cytoskeleton, altering the phosphorylation of macromolecules, etc. [65]. Additionally, cell death associated with prolonged convulsions can result in reactive gliosis. Taken together, these alterations can cause brain damage and persistent hyperstimulation. The mechanism of seizure induction for the KA model is very similar to the PILO model. Experimental evidence indicates that KA causes cell death in the hippocampus, the entorhinal cortex and the medial thalamic nuclei [5, 6, 64]. The kindling model involves the repeated application of a subconvulsive stimulus delivered through a bipolar electrode implanted into a limbic structure, such as the amygdala, hippocampus or entorhinal cortex [2]. Pitkanen and Sutula [52] reported neuronal damage, gliosis, neurogenesis and mossy fiber sprouting after using the kindling model of epilepsy.

## Neurogenesis: basic and new findings

As mentioned in the Introduction, neurogenesis takes place in specific brain regions throughout life. These regions are the subgranular zone (SGZ) of the DG in the hippocampus and the subventricular zone (SVZ) of the anterior lateral ventricles [21, 22, 44, 72]. According to recent data, newly born neurons from the SGZ are functionally integrated into the hippocampal circuitry [53, 79], and they have passive membrane properties, action potentials, and functional synaptic inputs similar to those found in mature granule cells [78]. The highest level of neurogenesis within the neurogenic zones occurs during prenatal development. However, generation of new neurons in the adult hippocampus has been one of the most provocative research topics for the last several years. The interest in neurogenesis in adolescence and adulthood increased rapidly because of the importance of hippocampal function in learning and memory in both the normal and the diseased/injured brain [40]. Alterations in cognitive function are commonly observed in TLE, Alzheimer's disease, cranial irradiation and traumatic brain injury [1, 16, 17, 54, 56]. Additionally, methods currently used to treat brain disorders can also significantly impact cognitive functions [14, 66].

#### Neurogenesis and acute seizures

Animal studies revealed that there are many conditions that affect the rate of cell proliferation, migration and differentiation and hence affect the process of neurogenesis. These changes are usually caused by environmental and pathological conditions. One of the most common abnormal conditions that has a significant impact on neurogenesis is a seizure. Many different studies have been performed to identify the disturbances in neurogenesis based on the types of convulsions (Tab. 1). Moreover, the same methods producing seizures can produce different types of neurogenesis alterations.

Electrical stimulation increases the rate of neurogenesis by neuronal depolarization or repetitive dis-

 Tab. 1. Neurogenesis after acute seizures (in vivo studies)

Type of seizure	Neurogenesis	Reference
KA (rats)	$\uparrow$	[43]
KA (rats)	$\uparrow$	[10]
Electrolytic lesions (rats)	$\uparrow$	[23]
Kindling (rats)	$\uparrow$	[7]
KA (rats)	$\uparrow$	[26]
PILO (rats)	$\uparrow$	[15]
Kindling (rats)	$\uparrow$	[4]
PILO (mice)	$\uparrow$	[33]

↑ increase in neurogenesis, KA = kainic acid, PILO = pilocarpine

charge [7]. Dramatic increases in neurogenesis were observed in the SGZ following PILO- and KAinduced SE [7, 26, 50] or kindling stimulation. Examination of the hippocampus from young TLE patients (2 years old) also suggested increased cell proliferation [8]. Although the molecular mechanisms underlying the seizure-induced increase in neurogenesis are unclear, several potential mechanisms have been proposed:

1. Up-regulation of the factors promoting proliferation and survival of neurons (including NGF – nerve growth factor, BDNF – brain-derived neurotrophic factor, FGF-2 – fibroblast growth factor, and VEGF – vascular endothelial growth factor) [4, 10, 15, 23, 25, 43].

2. Positive impact of increased levels of the inhibitory neurotransmitter  $\gamma$ -aminobutyric acid (GABA) on the proliferation of neural progenitors, the migration and differentiation of neuroblasts, and synaptic integration of newborn neurons in the DG a short time after a seizure [24].

3. Increase of neurogenesis in the presence of high levels of neuropeptide Y (NPY) [32, 33, 61, 62], which produces an increase in neural stem cell proliferation as a consequence of modulation of neuron-restrictive silencing factor (NRSF) activity [36].

Data from studies over the last several years show that the altered neurogenesis due to acute seizure activity has a strong impact on the development of aberrant circuitry [40]. Hippocampal injury in chronic epilepsy may occur as a consequence of acute seizures that disturb cell proliferation.

## Neurogenesis and chronic TLE

In rats, the increased neurogenesis observed after acute seizures usually returns to basal levels within 2 months after the convulsions [18]. However, in chronic TLE, which is characterized by high numbers of spontaneous seizures, hippocampus-dependent learning and memory deficits are observed, which can be at least partially linked to decreased neurogenesis [28]. Significant evidence for reduced neurogenesis in the KA model of mouse TLE was shown by Kralic and coworkers [39], who demonstrated a correlation between decreased neurogenesis and increased astrocyte production. Additional studies performed by Lederberger and colleagues [42] using the KA mouse model of TLE showed impaired fate commitment and/or early differentiation of proliferating cells in the lesioned DG.

The number of spontaneous seizures has a strong impact on the extent of neurogenesis. Significant alterations in the proliferation of neural progenitors and the migration and differentiation of neuroblasts were strongly associated with high numbers of spontaneous seizures. Heinrich and coworkers [30] showed a decrease in neurogenesis at 1 week and a virtual loss of all neurogenesis by 4–6 weeks after the first seizures. However, the opposite results were shown by Bonde et al. [9] in the electrically evoked SE rat model: no changes in neurogenesis (up to 6 months after SE) were reported. Moreover, no effects on neurogenesis were observed by Cha et al. [13], who found that the hippocampus was capable of generating new neurons several weeks after SE and that recurrent seizures enhanced the production of new neurons in a rat PILO model of epilepsy. Hattiangady and coworkers [29] showed that in the rat, severely diminished dentate gyrus neurogenesis in chronic TLE was not associated with either decreased production of new cells or reduced survival of newly born cells in the subgranular zone and in the granular cell layer. Rather, it was linked to a dramatic decline in the neuronal fate choice of newly generated cells. In chronic TLE, newly born cells differentiated primarily into glial cells, which was different from the neuronal fate that was observed in normal animals.

Waldau and colleagues [76] tested the hypothesis that spontaneous recurrent motor seizures (SRMS) in chronic TLE could be reduced by grafting neural stem cells that are capable of adding new GABA-ergic interneurons and glial-derived neurotrophic factor-ex-

Tab. 2. Neurogenesis after chronic spontaneous seizures (*in vivo* studies)

Type of seizure	Neurogenesis	Reference
KA (rats) PILO (rats) KA (mice) KA (mice) Electrical stimulation (rats) KA (rats)	↓ ↓ ↓ ↓ ↓ ↓ ↓ ↓ ↓	[28] [13] [39] [42] [30] [9] [29]

 $\downarrow$  decrease in neurogenesis,  $\uparrow$  increase in neurogenesis, KA = kainic acid, PILO = pilocarpine

pressing astrocytes into the epileptic hippocampus. They showed that the grafted neural stem cells decreased the number of spontaneous recurrent motor seizures in the rat model of chronic TLE. SRMS were reduced by 43%, and stage V seizures were reduced by 90%. This evidence confirms that this therapy effectively diminishes SRMS in chronic TLE. Based on the results obtained from different animal models of epilepsy, we can confirm that decreased neurogenesis depends on the animal model and on the age of the animal at the time of the initial seizure episode (Tab. 2).

# Neurogenesis after administration of antiepileptic drugs

Nearly 30% of epilepsy patients suffer from refractory epilepsy and require at least two antiepileptic drugs (AEDs) in combination. Each additional AED added to the standard treatment increases the risk of serious side effects. One very important side effect may be the alteration of neurogenesis. This can be very problematic, especially for children, because neurogenesis is most vigorous at early ages and because any reduction may cause learning and memory impairments. Treatment with AEDs takes at least 2-3 years and, in many situations, lasts a lifetime. While the most desirable effect is the reduction of seizures, long-term medication with AEDs may affect neuronal excitability and impair cognition and memory [49]. Reduced neurogenesis is often associated with alterations in some of the hippocampus-dependent cognitive functions [34, 55]. However, seizures are known to stimulate neurogenesis [37, 50]. This is why the problem of long-term treatment of epilepsy patients with AEDs is an important issue with respect to neurogenesis.

Results obtained by Chen et al. [14] using topiramate and lamotrigine, two commonly applied secondgeneration AEDs, indicated that topiramate but not lamotrigine promoted aberrant neuron regeneration in the hippocampus after SE. Similarly to lamotrigine, levetiracetam suppressed the development of spontaneous electroencephalographic (EEG) seizures and aberrant neurogenesis following KA-induced SE [71]. Moreover, Jessberger and coworkers showed that valproic acid (VPA – classical antiepileptic drug and mood stabilizer) potently blocked seizure-induced neurogenesis, an effect that appeared to be mainly mediated by inhibiting histone deacetylases (HDAC) and normalizing HDAC-dependent gene expression within the epileptic dentate area [36]. Additional studies performed on VPA indicated that it reduced cell proliferation in the dentate SGZ and impaired the ability of treated rats to successfully perform a hippocampus-dependent spatial memory test [75]. Moreover, the NMDA antagonist MK801 and two GABA<sub>A</sub> agonists, phenobarbital and diazepam, reduced numbers of newly born neurons in the brains of infant rats [70]. In the DG, many of the newly formed cells differentiated toward a neuronal phenotype; phenobarbital and MK801 significantly reduced the number of new neurons in that structure [70]. These results showed that NMDA receptor- and GABA<sub>A</sub> receptormediated enhancement disturbed cell proliferation and, in fact, inhibited neurogenesis.

## Conclusions

There is no doubt that adult hippocampal neurogenesis is very important for proper learning and memory. It is obvious that defective neurogenesis may contribute to progressive memory dysfunction [31]. The most popular animal model of TLE is widely used to better understand the process of epileptogenesis and to find the best way to protect neurons. It is very important for patients suffering from epilepsy to receive a medication that will be able to stop the process of epileptogenesis and protect neurons with minimal side effects, especially learning and memory disturbances. Results from animal models show that at early time points after acute seizures or SE, hippocampal neurogenesis and abnormal recruitment of newly born neurons into hippocampal circuitry increases, whereas the chronic phase of epilepsy is associated with substantially decreased hippocampal neurogenesis.

It is difficult to unambiguously determine if the reaction of increased neurogenesis after seizures increases or decreases the likelihood of epileptogenesis. However, recent studies have shown that seizureinduced neurogenesis has a pro-epileptogenic role in the formation of the epileptic hippocampus. The process of epileptogenesis generates ectopic granule cells that are born after SE and, instead of migrating to the granule cell layer, migrate to the hilus or to the inner molecular layer of the DG [48]. This situation is abnormal and may be one of the stimuli that induce epileptogenesis. Seizure activity or high levels of inflammation, which are main epileptogenic factors, have a strong impact on plasticity, migration patterns, morphology and the afferent synapses of the newly born cells. To answer the question of whether or not seizure-induced neurogenesis increases or decreases the likelihood of epileptogenesis, more advanced studies are necessary. These studies should focus on the integration of newly born cells into the existing neuronal network and on how this integration contributes to the excitability of this network.

Determining the best combination of AEDs is equally important because they can have a positive impact on neurogenesis. Therefore, studies concerning the problems of increased neurogenesis in TLE, learning and memory impairments and mood disorders are essential to better understand seizure development and to determine how to stop the process of epileptogenesis.

### **References:**

- Andres-Mach M, Rola R, Fike JR: Radiation effects on neural precursor cells in dentate gyrus. Cell Tissue Res, 2008, 331, 251–262.
- 2. Arida RM, Scorza FA, Scorza CA, Cavalheiro EA: Is physical activity beneficial for recovery in temporal lobe epilepsy? Evidences from animal studies. Neurosci Biobehav Rev, 2009, 33, 422–431.
- Avanzini G, Mosche SL, Schwartzkroin PA, Engel J: Animal models of localization–related epilepsy. In: Epilepsy: a Comprehensive Textbook. Eds. Engel J, Pedley TA, Philadelphia, Lippincott-Raven, 1997, 37, 427–442.
- Banerjee SB, Rajendran R, Dias BG, Ladiwala U, Tole S, Vaidya VA: Recruitment of the Sonic hedgehog signalling cascade in electroconvulsive seizure-mediated regulation of adult rat hippocampal neurogenesis. Eur J Neurosci, 2005, 22, 1570–1580.
- Ben-Ari Y, Tremblay E, Ottersen OP: Injections of kainic acid into amygdaloid complex of the rat: an electrographic, clinical and histological study in relation to the pathology of epilepsy. Neuroscience, 1980, 5, 515–528.
- Ben-Ari Y, Tremblay E, Ottersen OP, Meldrum BS: The role of epileptic activity in hippocampal and remote cerebral lesions induced by kainic acid. Brain Res, 1980, 191, 79–97.
- Bengzon J, Kokaia Z, Elmér E, Nanobashvili A, Kokaia M, Lindvall O: Apoptosis and proliferation of dentate gyrus neurons after single and intermittent limbic seizures. Proc Natl Acad Sci USA, 1997, 94, 10432–1047.
- Blumcke I, Schewe JC, Normann S, Brüstle O, Schramm J, Elger CE, Wiestler OD: Increase of nestin-immunoreactive neural precursor cells in the dentate gyrus of pediatric

patients with early-onset temporal lobe epilepsy. Hippocampus, 2001, 11, 311–321.

- Bonde S, Ekdahl CT, Lindvall O: Long-term neuronal replacement in adult rat hippocampus after status epilepticus despite chronic inflammation. Eur J Neurosci, 2006, 23, 965–974.
- Bugra K, Pollard H, Charton G, Moreau J, Ben-Ari Y, Khrestchatisky M: aFGF, bFGF and flg mRNAs show distinct patterns of induction in the hippocampus following kainate-induced seizures. Eur J Neurosci, 1994, 6, 58–66.
- 11. Cavalheiro EA: The pilocarpine model of epilepsy. Ital J Neuro Sci, 1995, 16, 33–37.
- Cavalheiro EA, Leite JP, Bortolotto ZA, Turski WA, Ikonomidou C, Turski L: Long-term effects of pilocarpine in rats: structural damage of the brain triggers kindling and spontaneous recurrent seizures. Epilepsia, 1991, 32, 778–782.
- Cha BH, Akman C, Silveira DC. Liu X, Holmes GL: Spontaneous recurrent seizure following status epilepticus enhances dentate gyrus neurogenesis. Brain Dev, 2004, 26, 394–397.
- Chen J, Quan QY, Yang F, Wang Y, Wang JC, Zhao G, Jiang W: Effects of lamotrigine and topiramate on hippocampal neurogenesis in experimental temporal-lobe epilepsy. Brain Res, 2010, 1313, 270–282.
- Croll SD, Goodman JH, Scharfman HE: Vascular endothelial growth factor (VEGF) in seizures: a double-edged sword. Adv Exp Med Biol, 2004, 548, 57–68.
- Detour J, Schroeder H, Desor D, Nehlig A: A 5-month period of epilepsy impairs spatial memory, decreases anxiety, but spares object recognition in the lithium-pilocarpine model in adult rats. Epilepsia, 2005, 46, 499–508.
- Devinsky O: Diagnosis and treatment of temporal lobe epilepsy. Rev Neurol, 2004, 1, 2–9.
- Deisseroth K, Singla S, Toda H, Monje M, Palmer TD, Malenka RC: Excitation-neurogenesis coupling in adult neural stem/progenitor cells. Neuron, 2004, 42, 535–552.
- Doetsch F, Garcia-Verdugo JM, Alvarez-Buylla A: Cellular composition and three-dimensional organization of the subventricular germinal zone in the adult mammalian brain. J Neurosci, 1997, 17, 5046–5061.
- Eriksson PS, Perfilieva E, Bjork-Eriksson T, Alborn AM, Nordborg C, Peterson DA: Neurogenesis in the adult human hippocampus. Nat Med, 1998, 4, 1313–1317.
- 21. Gage FH: Mammalian neural stem cells. Science, 2000, 287, 1433–1438.
- Gage FH Ray J, Fisher LJ: Isolation, characterization, and use of stem cells from the CNS. Annu Rev Neurosci, 1995, 18, 159–192.
- Gall CM, Berschauer R, Isackson PJ: Seizures increase basic fibroblast growth factor mRNA in adult rat forebrain neurons and glia. Brain Res Mol Brain Res, 1994, 21, 190–205.
- 24. Ge S, Pradhan DA, Ming GL Song H: GABA sets the tempo for activity-dependent adult neurogenesis. Trends Neurosci, 2007, 30, 1–8.
- Gomez-Pinilla F, van der Wal EA, Cotman CW: Possible coordinated gene expressions for FGF receptor, FGF-5, and FGF-2 following seizures. Exp Neurol, 1995, 133, 164–174.

- Gray WP, Sundstrom LE: Kainic acid increases the proliferation of granule cell progenitors in the dentate gyrus of the adult rat. Brain Res, 1998, 790, 52–59.
- Hastings NB, Gould E: Rapid extension of axons into the CA3 region by adult-generated granule cells. J Comp Neurol, 1999, 413, 146–154. Erratum in: J Comp Neurol, 1999, 415, 4.
- Hattiangady B, Rao MS, Shetty AK: Chronic temporal lobe epilepsy is associated with severely declined dentate neurogenesis in the adult hippocampus. Neurobiol Dis, 2004, 17, 473–490.
- 29. Hattiangady B, Shetty AK: Decreased neuronal differentiation of newly generated cells underlies reduced hippocampal neurogenesis in chronic temporal lobe epilepsy. Hippocampus, 2010, 20, 97–112.
- 30. Heinrich C, Nitta N, Flubacher A, Müller M, Fahrner A, Kirsch M, Freiman T et al.: Reelin deficiency and displacement of mature neurons, but not neurogenesis underlie the formation of granule cell dispersion in the epileptic hippocampus. J Neurosci, 2006, 26, 4701–4713.
- Helmstaedter C: Effects of chronic epilepsy on declarative memory system. Prog Brain Res, 2002, 135, 439–453.
- Howell OW, Scharfman HE, Herzog H, Sundstrom LE, Beck-Sickinger A, Gray WP: Neuropeptide Y is neuroproliferative for post-natal hippocampal precursor cells. J Neurochem, 2003, 86, 646–659.
- 33. Howell OW, Silva S, Scharfman HE, Sosunov AA, Zaben M, Shatya A, McKhann G 2<sup>nd</sup> et al.: Neuropeptide Y is important for basal and seizure-induced precursor cell proliferation in the hippocampus. Neurobiol Dis, 2007, 26, 174–188.
- 34. Iamayoshi I, Sakamoto M, Ohtsuka T, Takao K, Miyakawa T, Yamaguchi M, Mori K et al.: Roles of continuous neurogenesis in the structural and functional integrity of the adult forebrain. Nat Neurosci, 2008, 11, 1153–1161.
- Jessberger S, Kempermann G: Adult-born hippocampal neurons mature into activity-dependent responsiveness. Eur J Neurosci, 2003, 18, 2707–2712.
- 36. Jessberger S, Nakashima K, Clemenson GD, Mejia E, Mathews E, Ure K, Ogawa S et al.: Epigenetic modulation of seizure-induced neurogenesis and cognitive decline. J Neurosci, 2007, 27, 5967–5975.
- 37. Jiang W, Wan Q, Zhang ZJ, Wang WD, Huang YG, Rao ZR, Zhang X: Dentate granule cell neurogenesis after seizures induced by pentylenetetrazol in rats. Brain Res, 2003, 977, 141–148.
- Kaplan MS, Bell DH: Mitotic neuroblasts in the 9-dayold and 11-month-old rodent hippocampus. J Neurosci, 1984, 4, 1429–1441.
- 39. Kralic JE, Ledergerber DA, Fritschy JM: Disruption of the neurogenic potential of the dentate gyrus in a mouse model of temporal lobe epilepsy with focal seizures. Eur J Neurosci, 2005, 22, 1916–1927.
- Kuruba R, Hattiangady B, Shetty AK: Hippocampal neurogenesis and neural stem cells in temporal lobe epilepsy. Epilepsy Behav, 2009, 14, 65–73.
- Kwan P, Sander JW: The natural history of epilepsy: an epidemiological view. J Neurol Neurosurg Psychiatry, 2004, 75, 1376–1381.

- Ledergerber D, Fritschy JM, Kralic JE: Impairment of dentate gyrus neuronal progenitor cell differentiation in a mouse model of temporal lobe epilepsy Exp Neurol, 2006, 199, 130–142.
- Lowenstein DH, Seren MS, Longo FM: Prolonged increases in neurotrophic activity associated with kainicinduced hippocampal synaptic reorganization. Neuroscience, 1993, 56, 597–604.
- 44. Luskin MB: Restricted proliferation and migration of postnatally generated neurons derived from the forebrain subventricular zone. Neuron, 1993, 11, 173–189.
- Luskin MB, Mc Dermott K: Divergent lineages for oligodendrocytes and astrocytes originating in the neonatal forebrain subventricular zone. Glia, 1994, 11, 211–226.
- 46. Łuszczki JJ, Jaskólska A, Dworzański W, Żółkowska D: 7-Nitroindazole, but not N<sup>G</sup>-nitro-L-arginine, enhances the anticonvulsant activity of pregabalin in the mouse maximal electroshock-induced seizure model. Pharmacol Rep, 2011, 63, 169–175.
- Mody I, Schwartzkroin PA: Acute seizure models (intact animals). In: Epilepsy: a Comprehensive Textbook. Eds. Engel J, Pedley TA, Philadelphia, Lippincott-Raven, 1997, 397–404.
- Morschead CM, Reynolds BA, Craig CG, McBurney MW, Staines WA, Morassutti D, van der Weiss S, Kooy D: Neural stem cells in the adult mammalian forebrain: a relatively quiescent subpopulation of subependymal cells. Neuron, 1994, 13, 1071–1082.
- Ortinsky P, Meador KJ: Neuronal mechanism of conscious awareness. Arch Neurol, 2004, 61, 1017–1020.
- Parent JM, Yu TW, Leibowitz RT, Geschwind DH, Sloviter RS, Lowenstein DH: Dentate granule cell neurogenesis is increased by seizures and contributes to aberrant network reorganization in the adult rat hippocampus. J Neurosci, 1997, 17, 3727–3738.
- Pierce JP, Melton J, Punsoni M, McCloskey DP, Scharfman HE: Mossy fibers are the primary source of afferent input to ectopic granule cells that are born after pilocarpine-induced seizures. Exp Neurol, 2005, 196, 316–331.
- 52. Pitkanen A, Sutula TP: Is epilepsy a progressive disorder? Prospects for new therapeutic approaches in temporal lobe epilepsy. Lancet Neurol, 2002, 1, 173–181.
- Ramirez-Amaya V, Marrone DF, Gage FH, Worley PF, Barnes CA: Integration of new neurons into functional neural networks. J. Neurosci, 2006, 26, 12237–12241.
- Rola R, Mizumatsu S, Otsuka S, Morhardt DR, Noble-Haeusslein LJ, Fishman K, Potts MB, Fike JR: Alterations in hippocampal neurogenesis following traumatic brain injury in mice. Exp Neurol, 2006, 202,189–199.
- 55. Rola R, Raber J, Rizk A, Otsuka S, VandenBerg SR, Morhardt DR, Fike JR: Radiation-induced impairment of hippocampal neurogenesis is associated with cognitive deficits in young mice. Exp Neurol, 2004, 188, 316–330.
- Rosi S, Andres-Mach M, Fishman KM, Levy W, Ferguson RA, Fike JR: Cranial irradiation alters the behaviorally induced immediate-early gene Arc (activity-regulated cytoskeleton-associated protein). Cancer Res, 2008, 68, 9763–9770.
- 57. Sander JW: Some aspects of prognosis in the epilepsies: a review. Epilepsia, 1993, 34, 1007–1016.

- 58. Sander JW: The epidemiology of epilepsy revisited. Curr Opin Neurol, 2003, 16, 165–170.
- Sarkisian MR: Overview of the current animal models of human seizure and epileptic disorders. Epilepsy Behav, 2001, 2, 201–216.
- 60. Scharfman HE, Goodman JH, Sollas AL: Granule-like neurons at the hilar/CA3 border after status epilepticus and their synchrony with area CA3 pyramidal cells: functional implications of seizure-induced neurogenesis. J Neurosci, 2000, 20, 6144–6158.
- Scharfman HE, Gray WP: Plasticity of neuropeptide Y in the dentate gyrus after seizures, and its relevance to seizure-induced neurogenesis. EXS, 2006, 95, 193–211.
- 62. Scharfman HE, Gray WP: Relevance of seizure-induced neurogenesis in animal models of epilepsy to the etiology of temporal lobe epilepsy. Epilepsia, 2007, 48, 33–41.
- 63. Schmidt-Hieber C, Jonas P, Bischofberger J: Enhanced synaptic plasticity in newly generated granule cells of the adult hippocampus. Nature, 2004, 429, 184–187.
- 64. Schwob JE. Fuller T, Price JL, Olney JW: Widespread patterns of neuronal damage following systemic or intracerebral injection of kainic acid: a histological study. Neuroscience, 1980, 5, 991–1014.
- 65. Scorza FA, Arida RM, Naffah-Mazzacorati MG, Scerni DA, Calderazzo L Cavalheiro EA: The pilocarpine model of epilepsy: what have we learned? An Acad Bras Cienc, 2009, 81, 345–365.
- Shatskikh T, Zhao Q, Zhou JL, Holmes GL: Effect of topiramate on cognitive function and single units from hippocampal place cells following status epilepticus. Epilepsy Behav, 2009, 14, 40–47.
- Shors TJ: Memory traces of trace memories: neurogenesis, synaptogenesis and awareness. Trends Neurosci, 2004, 27, 250–256.
- Shibley H, Smith BN: Pilocarpine-induced status epilepticus results in mossy fiber sprouting and spontaneous seizures in C57BL/6 and CD-1 mice. Epilepsy Res, 2002, 49, 109–120.
- Snyder JS, Kee N, Wojtowicz JM: Effects of adult neurogenesis on synaptic plasticity in the rat dentate gyrus. J Neurophysiol, 2001, 85, 2423–2431.
- Stefovska VG, Uckermann O, Czuczwar M, Smitka M, Czuczwar P, Kis J, Kaindl AM et al.: Sedative and anticonvulsant drugs suppress postnatal neurogenesis. Ann Neurol, 2008, 64, 434–445.
- Sugaya Y, Maru E, Kudo K, Shibasaki T, Kato N: Levetiracetam suppresses development of spontaneous EEG seizures and aberrant neurogenesis following kainate-induced status epilepticus. Brain Res, 2010, 1352, 187–199.
- Temple S, Alvarez-Buylla A: Stem cells in the adult mammalian central nervous system. Curr Opin Neurobiol, 1999, 9, 135–141.
- Turski WA, Cavalheiro EA, Bortolotto ZA, Mello LM, Schwarz M, Turski L: Seizures produced by pilocarpine in mice: a behavioral, electroencephalographic and morphological analysis. Brain Res, 1984, 321, 237–253.
- 74. Turski WA, Cavalheiro EA, Schwarz M, Czuczwar SJ, Kleinrok Z, Turski L: Limbic seizures produced by pilocarpine in rats: a behavioral, electroencephalographic

and neuropathological study. Behav Brain Res, 1983, 9, 315–335.

- Umka J, Mustafa S, ElBeltagy M, Thorpe A, Latif L, Bennett G, Wigmore PM: Valproic acid reduces spatial working memory and cell proliferation in the hippocampus. Neuroscience, 2010, 166, 15–22.
- 76. Waldau B, Hattiangady B, Kuruba R, Shetty AK: Medial ganglionic eminence-derived neural stem cell grafts ease spontaneous seizures and restore GDNF expression in a rat model of chronic temporal lobe epilepsy. Stem Cells, 2010, 28, 1153–1164.
- van Praag H, Schinder AF, Christie BR, Toni N, Palmer TD, Gage FH: Functional neurogenesis in the adult hippocampus. Nature, 2002, 415, 1030–1034.
- 78. van Praag H, Christie BR, Sejnowski TJ, Gage FH: Running enhances neurogenesis, learning, and long-term potentiation in mice. Proc Natl Acad Sci USA, 1999, 96, 13427–13431.
- Zhao C, Teng EM, Summers RG Jr, Ming GL, Gage FH: Distinct morphological stages of dentate granule neuron maturation in the adult mouse hippocampus. J Neurosci, 2006, 26, 3–11.

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