



Review

Novel approaches to improving endothelium-dependent nitric oxide-mediated vasodilatation

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Abstract:

Endothelial dysfunction, which is defined by decreased endothelium-dependent vasodilatation, is associated with an increased number of cardiovascular events. Nitric oxide (NO) bioavailability is reduced by altered endothelial signal transduction or increased formation of radical oxygen species reacting with NO. Endothelial dysfunction is therapeutically reversible and physical exercise, calcium channel blockers, angiotensin converting enzyme inhibitors, and angiotensin receptor antagonists improve flow-evoked endothelium-dependent vasodilatation in patients with hypertension and diabetes. We have investigated three different approaches, with the aim of correcting endothelial dysfunction in cardiovascular disease. Thus, (1) we evaluated the effect of a cell permeable superoxide dismutase mimetic, tempol, on endothelial dysfunction in small arteries exposed to high pressure, (2) investigated the endothelial signal transduction pathways involved in vasorelaxation and NO release induced by an olive oil component, oleanolic acid, and (3) investigated the role of calcium-activated K channels in the release of NO induced by receptor activation. Tempol increases endothelium-dependent vasodilatation in arteries from hypertensive animals most likely through the lowering of radical oxygen species, but other mechanisms also appear to contribute to the effect. While oleanolic acid leads to the release of NO by calcium-independent phosphorylation of endothelial NO synthase, endothelial calcium-activated K channels and an influx of calcium play an important role in G-protein coupled receptor-evoked release of NO. Thus, all three approaches increase bioavailability of NO in the vascular wall, but it remains to be addressed whether these actions have any direct benefit at a clinical level.

Key words: endothelium, nitric oxide, oleanolic acid, tempol, calcium-activated K channels

Abbreviations: BK_{Ca} – large-conductance calcium-activated K channels, EDHF – endothelium-derived hyperpolarizing factor, NO – nitric oxide, eNOS – endothelial NO synthase, O₂⁻ – superoxide anion, SOD – superoxide dismutase

Introduction

The endothelium regulates tone at rest and during exercise, the thrombotic and adhesive properties of the

vascular wall, the architecture of the vascular wall, and vascular permeability. Disturbances of these principal endothelial functions are termed endothelial dysfunction. Endothelium-dependent vasodilatation induced by increased blood flow and receptor-specific agonists, such as acetylcholine and bradykinin, are reduced in the presence of classical vascular risk factors including hypertension, hypercholesterolemia, diabetes, smoking, aging and by atherosclerotic disease and inflammation. Moreover, endothelial dysfunction in the coronary circulation [6, 63, 64, 80, 94] and the forearm [72] is associated with progression of ischemic heart disease, and endothelial dysfunction in the microvasculature is considered to be an independent risk factor for ischemic heart disease [43]. Therefore, disturbed endothelial function is of prognostic significance.

Early studies have suggested that endothelial dysfunction may be therapeutically reversible [17], and physical exercise, calcium channel blockers and angiotensin converting enzyme inhibitors and angiotensin receptor antagonists were found to improve flow-evoked endothelium-dependent vasodilation in patients with hypertension and diabetes [18, 40, 71, 81]. It is controversial whether lipid-lowering statins improve endothelium-dependent vasodilation in patients [18, 19, 96, 100] or have a minor or no effect on endothelium-dependent vasodilation [8, 30, 35, 103]. The latter can be ascribed to different clinical designs (see [44]), but also the potency, structure, and pleiotropic effects of each statin [67], which requires further investigation. However, correction of endothelial dysfunction in hypertension was found to have prognostic significance [58], hence supporting that there is an unmet need for drugs that specifically improve endothelial function.

A key mechanism underlying endothelial dysfunction and increased expression of adhesion molecules and chemoattractants is the loss of endothelial nitric oxide (NO) bioavailability. NO bioavailability is decreased either by decreased formation or by enhanced removal of NO. The presence of classical cardiovascular risk factors is associated with enhanced generation of radical oxygen species where superoxide anions (O_2^-) play a pivotal role by reacting with NO resulting in the formation of peroxynitrite ($ONOO^-$), and hence decreasing bioavailability of NO (see [15]). Moreover, we have found, by simultaneous measurements of relaxation and release of NO that endothelial cell signal transduction is altered in hypertension

[92]. Some of the specific treatments of endothelial dysfunction have recently been reviewed, including endothelin receptor antagonists [48], L-arginine [11], tetrahydrobiopterin analogues [3], 1-methylnicotinamide [5] and phosphodiesterase type 5 inhibitors (e.g., sildenafil and tadalafil) [87] (Fig. 1). The following is a review of three novel approaches that we have investigated with the aim of correcting endothelial dysfunction in cardiovascular disease (Fig. 2). Thus, (1) we evaluated the effect of a cell permeable superoxide dismutase mimetic, tempol, on endothelial dysfunction in small arteries exposed to high pres-

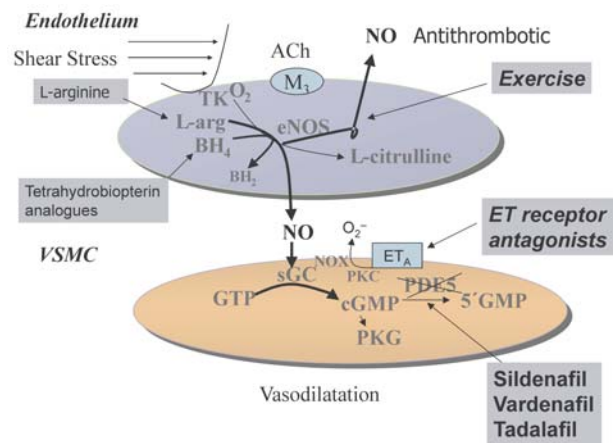


Fig. 1. Specific treatments to restore endothelium-dependent vasodilatation (grey boxes). sGC – soluble guanylyl cyclase; VSMC – vascular smooth muscle cell

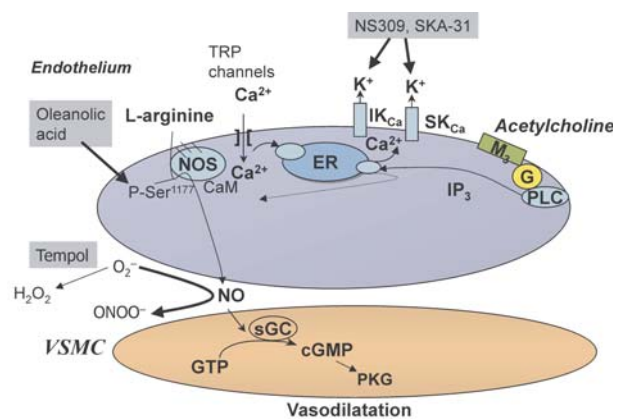


Fig. 2. Novel approaches to improve endothelium-dependent nitric oxide-mediated vasodilatation (grey boxes). SK_{Ca} – small conductance calcium-activated K channels; IK_{Ca} – intermediate calcium-activated K channels; sGC – soluble guanylyl cyclase; TRP – transient receptor potential channels; VSMC – vascular smooth muscle cell

sure, (2) investigated the endothelial signal transduction pathways involved in vasorelaxation and NO release induced by an olive oil component, oleanolic acid, and (3) investigated the role of calcium-activated K channels in the release of NO induced by receptor activation.

The superoxide dismutase mimetic tempol and endothelial dysfunction

The level of radical oxygen species is determined by the radical oxygen species generating enzymes in the extracellular and intracellular compartments depending on location, e.g., NAD(P)H/NADH oxidase in the cell membrane, xanthine oxidase, myeloperoxidase, cytochrome P450 in cytoplasm, and mitochondrial oxidation and antioxidant enzymes e.g., superoxide dismutases (SODs), glutathione peroxidase, heme oxygenase, thioredoxin peroxidase/peroxiredoxin, catalase, and paraoxonase [42]. The increased radical oxygen species levels are further enhanced by uncoupling of endothelial eNOS, which ceases to produce NO and instead switches to superoxide production [49, 104]. Thus, there are many sources in the vascular wall leading to radical oxygen species generation, making scavenging of radical oxygen species an attractive therapeutic approach for treating endothelial dysfunction associated with cardiovascular disease.

There are natural extra- and intracellular antioxidant defense mechanisms that scavenge radical oxygen species. For example, SOD catalyzes the conversion of O_2^- to H_2O_2 , which can be turned into water by catalase or the glutathione peroxidase system. Although SOD is the first line of physiological defense against oxidative stress, the reaction of O_2^- with NO is about three times faster than its reaction with SOD [7]. However, the addition of an exogenous SOD mimetic, tempol, protects animals and mammalian cells from cytotoxicity induced by oxygen-free radicals like hydroxyl radicals, H_2O_2 , and O_2^- [57, 78]. An additional property of tempol as antioxidant is that it can penetrate cell membranes, and hence react with both intracellular and extracellular oxygen-free radicals. These properties make tempol attractive for treatment of cardiovascular disease associated with oxidative stress (e.g., hypertension).

In rat small mesenteric arteries activated with a thromboxane analogue, increases in flow and acetylcholine induce endothelium-dependent vasodilatation, which, in the case of flow, appears to be mediated by NO, while acetylcholine evokes mainly endothelium-derived hyperpolarizing factor (EDHF) type relaxations [98]. Exposing isolated pressurized arteries to elevated pressure for one hour and then returning intraluminal pressure to normotensive levels results in impaired vasodilation in response to flow, while acetylcholine-evoked vasodilatation is conserved [24]. These findings suggest that bioavailability of NO is selectively decreased by high pressure and underlies impaired flow-mediated vasodilatation. Our findings are in line with those from previous studies in models of hypertension *in vivo* [36, 110] and in rat isolated skeletal arterioles [101], where elevated pressure was also shown to elicit arterial O_2^- production by activation of NADPH oxidase resulting in impaired endothelial function. Moreover, these results were further supported by the observations that vasodilatation to an NO donor, S-nitroso-N-acetylpenicillamine was also reduced, while vasodilation in response to a direct guanylyl cyclase activator, BAY 412272, were conserved, and O_2^- formation was increased in arterial segments exposed to high pressure in small mesenteric arteries [24]. These results suggest that O_2^- , by reaction with NO, reduces NO bioavailability and leads to impaired flow-induced NO-mediated dilatation in arteries exposed to high pressure; indeed, in the *in vitro* model of endothelial dysfunction in small arteries induced by elevated pressure, incubation of the SOD mimetic tempol conserved the vasodilatation most likely by lowering O_2^- levels in the arterial wall [24].

There have been numerous studies performed on arteries from different animal models showing that treatment or incubation with tempol restores agonist-induced endothelium-dependent vasodilatation. Thus, tempol restored acetylcholine and arachidonic acid relaxation in skeletal muscle arteries and coronary arteries from diabetic animals [38, 106], and acetylcholine relaxation in afferent arterioles from Dahl salt sensitive rats [65] and in carotid arteries from deoxycorticosterone acetate-salt hypertensive rats [65, 111]. That tempol can substitute for endogenous SOD is also supported by the observations that tempol treatment restores acetylcholine relaxation in carotid arteries from mice heterozygous for CuZn SOD [27, 108]. Therefore, to investigate whether tempol is also able

to restore endothelial function in small arteries exposed to high blood pressure *in vivo*, one-kidney one-clip hypertensive rats were treated with tempol [25]. In arteries from the renal hypertensive rats, flow-induced dilatation was blunted compared to normotensive and tempol-treated rats, while acetylcholine-induced dilatation remained normal [25]. As measured by dihydroethidium staining, there was an increased level of O_2^- in arteries from vehicle-treated rats, but not from tempol-treated rats [25]. These studies suggest that renal hypertension, similar to elevated pressure *in vitro*, selectively inhibits flow-induced NO-mediated vasodilatation by increased generation of O_2^- contributing to decreased NO bioavailability. Furthermore, the endothelial dysfunction observed can be corrected by tempol treatment.

The findings of less O_2^- in the arterial wall of arteries from hypertensive rats treated with tempol suggest that tempol is a SOD mimetic, but do not exclude other mechanisms also contributing to the vasodilator effect. Thus, similar to endogenous SOD, tempol not only reduces O_2^- , but may also increase H_2O_2 in blood vessels and the kidney medulla [21, 22]. In mouse cremaster arterioles, tempol (1 mM) induced a transient and catalase-sensitive vasodilatation, suggesting it was mediated by H_2O_2 [21]. Moreover, in mesenteric arteries from Cu, Zn-SOD^{-/-} mice, tempol increased acetylcholine-induced catalase-sensitive vasodilatation and increased vascular H_2O_2 [108]. Although H_2O_2 can also induce vascular contraction [4, 39], these studies suggest H_2O_2 can contribute to the effects of tempol and that an enhanced vasodilatory effect by adding tempol should not only be attributed to dismutation of O_2^- anion and increased bioavailability of NO. Moreover, isolated patch clamp studies on vascular smooth muscle cells from normotensive and deoxycorticosterone acetate-salt hypertensive rats suggest that high tempol concentrations (~1 mM) increase a large-conductance calcium-activated potassium (BK_{Ca}) current [107]. Therefore, high concentrations of tempol appear to activate BK_{Ca} channels directly, whereas it remains to be addressed whether lower concentrations of NO, indirectly through formation of H_2O_2 or increased bioavailability, can lead to vasodilatation through activation of BK_{Ca} channels.

In humans, endothelial dysfunction is a marker of cardiovascular disease, and compromised flow-mediated dilatation is associated with a worse prognosis. In cross sectional studies, flow-mediated dilatation is linked with increased oxidative stress markers [95],

and a main contributor to a reduced flow-mediated dilatation is increased O_2^- production and reduced bioavailability of NO [61]. Thus, a reduction in O_2^- generation would be a logical therapeutic approach. However, it currently remains unproven that a reduction in the level of oxidative stress *per se* improves outcome of cardiovascular diseases in humans. Potential therapies for lowering of radical oxygen species include existing drug treatment with angiotensin-converting enzyme inhibitors [53], statins [23], and the β -adrenoceptor antagonists, carvedilol and nebivolol [2]. Dietary polyphenols present in wine, green tea, and fruits are mild antioxidants and may also offer long-term benefits by lowering radical oxygen species [68]. Moreover, vitamin C and vitamin E were found to lower oxidative stress, but large-scale clinical studies have not supported an effect of vitamin C or vitamin E supplementation on cardiovascular disease or mortality [10, 51]. However, the drugs that affect O_2^- levels in humans, such as inhibitors of the renin angiotensin aldosterone system, some beta-adrenoceptor antagonists and statins, like tempol, have additional effects including lowering of blood pressure or the cholesterol concentration, which by themselves lower O_2^- production. It is therefore important that tempol is compared to existing cardiovascular drugs. However, as tempol clearly has a different mechanism of action from these compounds, it is possible that a combination of tempol and conventional therapy could result in improved endothelial function and patient outcome.

The olive oil component, oleanolic acid, and endothelial dysfunction

Olive oil is an integral ingredient of the Mediterranean diet, and accumulating evidence suggests that it may have healthy benefits in terms of reduction of cardiovascular risk factors [32, 99]. The cardioprotective effects of olive oil have been ascribed to its content of monounsaturated fatty acids such as oleic acid and the presence of other biologically minor constituents such as polyphenols, tocopherols and triterpenoids [32, 54]. Olive pomace oil, so-called *orujo* olive oil in Spain, is obtained from the residue that remains after virgin olive oil has been mechanically extracted. As a consequence of these extraction processes, po-

mace olive oil differs from virgin olive oil in the content of minor components. In this regard, despite the lack of polyphenols, pomace olive oil contains higher concentrations of triterpenic compounds, such as oleanolic acid, than virgin olive oil [70]. Recent research has demonstrated that chronic parenteral administration of oleanolic acid prevents hypertension in salt-sensitive rats [88]. Furthermore, ingestion of pomace olive oil with a high proportion of oleanolic acid delays progression of lipid peroxidation in rat liver microsomes [69]. These studies suggest that oleanolic acid may contribute to the cardioprotective effect of ingestion of a diet rich in olive oil.

In aortic segments from normotensive and hypertensive rats, oleanolic acid evokes endothelium-dependent relaxations sensitive to inhibition of eNOS [74]. Investigation of large and small mesenteric arteries from rats also revealed that oleanolic acid induces endothelium-dependent relaxations sensitive to inhibition of NO synthase [76]. Moreover, inhibition of cyclooxygenase with indomethacin and of EDHF-type relaxation with a combination of calcium-activated K channel blockers, apamin and charybdotoxin, failed to alter oleanolic acid relaxation. These findings suggest that oleanolic acid evokes endothelium-dependent vasodilatation mediated by NO, and simultaneous measurements of NO and relaxation revealed that oleanolic acid evokes relaxation by release of NO [76].

Endothelial NO is produced by eNOS, which is classically activated by agonists such as acetylcholine, histamine and bradykinin inducing an increase in intracellular calcium levels followed by calcium/calmodulin-dependent modulation [59]. In contrast to acetylcholine, oleanolic acid failed to increase endothelial cell calcium, and hence evoked relaxation and release of NO independent of endothelial cell calcium in rat superior mesenteric arteries [76]. However, over the past five to 10 years, a far more complex model of eNOS regulation has emerged involving post-translational mechanisms such as protein-protein interactions and tightly regulated multisite phosphorylation. Overall, Ser¹¹⁷⁷ is the most extensively studied and appears to be the most important of the regulatory eNOS phosphorylation sites [34]. Phosphorylation of eNOS-Ser¹¹⁷⁷ is associated with an increase in activation and NO production in response to a growing list of stimuli, including mechanical factors (e.g., shear stress) [12] and humoral factors (e.g., insulin) [60] even at resting levels of calcium [55, 60]. Estrogen,

mechanical shear stress, and inhibitors of tyrosine phosphatase such as vanadate have been demonstrated to cause eNOS-Ser¹¹⁷⁷ activation and NO release by calcium-independent mechanisms [26, 66, 77]. eNOS-Ser¹¹⁷⁷ can be phosphorylated by several protein kinases, including Akt/protein kinase B, protein kinase A and AMP-activated protein kinase [12, 28, 37]. Therefore, in human umbilical cord endothelial cells, we investigated these pathways and found that oleanolic acid time-dependently increased phosphorylation of Akt kinase at Serine⁴⁷³ and eNOS at Serine¹¹⁷⁷, where this effect was blunted by inhibition of the upstream enzyme, phosphoinositide-3-kinase [76]. These findings suggest that oleanolic acid and acetylcholine increase vascular NO concentration by different signal transduction pathways, and that oleanolic acid by activation of phosphoinositide-3-kinase and phosphorylation of Akt can increase eNOS activity and NO production.

Several observations suggest that the effect of oleanolic acid on cell function is specific. Thus, structurally modified analogues of oleanolic acid bind with different potency to Kelch-like-ECH-associated protein 1, a cytoplasmic repressor of the transcription factor, NRF2 [29]. The structurally-related compounds, oleanolic acid, erythrodiol, maslinic acid, and uvaol, which only differ in substitution of one chemical group, cause relaxations with different potency and magnitude in rat aorta [29, 75], and caulophyllogenin, which is structurally related to oleanolic acid and only differs by a couple of hydroxyl substitutions, failed to induce relaxation in rat mesenteric arteries. Although these findings suggest the effects of oleanolic acid on endothelial cell function are specific, further studies are required to clarify the role of structural modifications of oleanolic acid for release of NO and endothelium-dependent relaxation as well as for the anti-inflammatory effects of the compound.

The clinical relevance of the NO releasing and relaxing activity of oleanolic acid depends on the systemic availability. In olive oil, the content of oleanolic acid is approximately 56 mg kg⁻¹ and in orujo olive oil it is 416 mg kg⁻¹ [73]. Recent studies reveal that, after ingestion of 40 mg oleanolic acid, plasma concentrations reached 12 ng ml⁻¹ [89], which is below the oleanolic acid concentrations causing release of NO and relaxation. That systemic availability of oleanolic acid is important is also supported by the observation that intraperitoneal administration of oleanolic acid lowers blood pressure in hypertensive

rats [88], but there is no effect on blood pressure of oleanolic acid administered in the diet [73]. However, in the same study, pomace olive oil supplemented in oleanolic acid increased eNOS expression and improved vasorelaxation in aortic segments from spontaneous hypertensive rats [73]. The latter findings support that systemic availability of oleanolic acid by administration in the diet is sufficient to improve endothelial function.

Oleanolic acid is a novel approach to improving endothelial function and may explain some of the beneficial effects on cardiovascular disease of a Mediterranean diet. However, oleanolic acid and structurally similar compounds should be tested in other animal models to determine whether they improve endothelial dysfunction induced by other cardiovascular risk factors, and it remains to be addressed whether the *in vivo* effects of oleanolic acid are due to an effect on eNOS activity and/or anti-inflammatory effects of the compound.

Therapeutic potential of targeting calcium-activated K channels in the endothelium

By simultaneous measurements of relaxation and release of NO in arteries from renal hypertensive rats, we have found that treatment with superoxide dismutase or L-arginine *per se* does not restore NO bioavailability [92]. Moreover, treatment of renal hypertensive animals with tempol restores vasorelaxation, but fails to restore NO release induced by acetylcholine in rat coronary arteries [25]. These findings suggest that endothelial cell signal transduction is also altered in hypertension, and that a therapeutic potential to treat endothelial dysfunction would be modulation of the endothelial signal transduction pathways.

The inhibition by apamin and charybdotoxin, blockers of small- and intermediate-conductance Ca^{2+} -activated K channels, has been considered a unique characteristic of EDHF type relaxation [14, 31, 109, 112]. Acetylcholine-evoked hyperpolarization of the endothelial cell coincides with rises in intracellular calcium in endothelial cells in intact rat aorta [16, 102], and patch clamp experiments and RT-PCR have provided evidence for the presence of small, intermediate and large-conductance Ca^{2+} -activated K channels in

endothelial cells of intact arteries [50, 52, 62]. Therefore, it was suggested that apamin and charybdotoxin may exert their effects mainly *via* Ca^{2+} -activated K channels located on the endothelial cells rather than on smooth muscle cells [9, 13, 31]. In the superior mesenteric artery, the combination of apamin and charybdotoxin also inhibits smooth muscle hyperpolarization and acetylcholine relaxation [20]. However, in contrast to small mesenteric arteries where acetylcholine mainly evokes EDHF type relaxations [47, 84, 98], acetylcholine evokes endothelium-dependent NO release and relaxation, which are blunted in the presence of a NOS inhibitor and abolished by combination with a NO scavenger, oxyhemoglobin, in rat superior mesenteric artery [86, 91, 92]. Therefore, we addressed whether agonist-evoked increases in NO concentration in intact segments of rat superior mesenteric arteries also involved activation of calcium-activated K channels of small and intermediate conductance. A combination of blockers of these channels, apamin and charybdotoxin markedly inhibited an acetylcholine-evoked increase in NO concentration and relaxation [91]. Despite causing contraction, noradrenaline also increases endothelial cell calcium and NO concentration [45], and a noradrenaline-evoked increase in NO concentration was also blunted in the presence of apamin and charybdotoxin [91]. These findings suggest that receptor-activated endothelial signal transduction leading to an increase in endothelial cell calcium is followed by activation of calcium-activated K channels, hyperpolarization and most likely amplified NO release.

Based on patch clamp studies and measurements of intracellular calcium in cultured endothelial cells, it has been suggested that the opening of calcium-activated K channels of small and intermediate conductance is coupled to increased calcium influx [82]. However, in intact arteries, acetylcholine and noradrenaline-evoked endothelial cell calcium levels remain unaltered in the presence of a combination apamin and charybdotoxin, which blocks these channels [41, 56, 76, 91]. Therefore, it remains to be addressed how opening of calcium-activated K channels of small and intermediate conductance are coupled to release of NO.

Knockout of endothelial calcium-activated K channels of small [97] and intermediate conductance [85] are associated with impaired endothelium-dependent vasodilatation and hypertension. The expression of small and intermediate conductance calcium-activated

K channels is reduced in angiotensin II-induced hypertension [46], while expression is unaltered in mesenteric arteries from diabetic rats, but the current is reduced [1]. Small molecules (NS309 and SKA31) modulating these channels are available [93, 105], and have recently been shown to induce hyperpolarization [1], release of NO in segments of rat mesenteric [90] and skeletal arteries [83], enhancement of receptor-activated EDHF-type relaxation in small arteries from mice [79], and induction of relaxation in human arteries [33] (Fig. 2). Therefore, given that opening of calcium-activated K channels of small and intermediate conductance are involved in both release of NO and EDHF-type relaxation, and that small molecules are available, provides these channels as attractive targets for exploring improvements in endothelium-dependent vasodilatation in cardiovascular disease.

Conclusion and perspectives

In summary, the three approaches that we have applied here show that: (1) tempol increases endothelium-dependent vasodilatation in arteries from hypertensive animals, most likely through a lowering of radical oxygen species, but that other mechanisms also appear to contribute to the effect, (2) oleanolic acid leads to the release of NO by calcium-independent phosphorylation of endothelial NO synthase, (3) endothelial calcium-activated K channels and influx of calcium play an important role in G-protein coupled receptor-evoked release of NO. Thus, all three approaches increase bioavailability of NO in the vascular wall, but it remains to be addressed whether they have any direct benefit at the clinical level.

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References:

1. Absi M, Burnham MP, Weston AH, Harno E, Rogers M, Edwards G: Effects of methyl beta-cyclodextrin on EDHF responses in pig and rat arteries; association between SK_{Ca} channels and caveolin-rich domains. *Br J Pharmacol*, 2007, 151, 332–340.
2. Agabiti RE, Rizzoni D: Metabolic profile of nebivolol, a beta-adrenoceptor antagonist with unique characteristics. *Drugs*, 2007, 67, 1097–1107.
3. Alp NJ, Channon KM: Regulation of endothelial nitric oxide synthase by tetrahydrobiopterin in vascular disease. *Arterioscler Thromb Vasc Biol*, 2004, 24, 413–420.
4. Ardanaz N, Beierwaltes WH, Pagano PJ: Distinct hydrogen peroxide-induced constriction in multiple mouse arteries: potential influence of vascular polarization. *Pharmacol Rep*, 2008, 60, 61–67.
5. Bartuś M, Łomnicka M, Kostogryś RB, Kaźmierczak P, Watała C, Słomińska EM, Smoleński RT et al.: 1-Methylnicotinamide (MNA) prevents endothelial dysfunction in hypertriglyceridemic and diabetic rats. *Pharmacol Rep*, 2008, 60, 127–138.
6. Bauersachs J, Widder JD: Endothelial dysfunction in heart failure. *Pharmacol Rep*, 2008, 60, 119–126.
7. Beckman JS, Koppenol WH: Nitric oxide, superoxide, and peroxynitrite: The good, the bad, and ugly. *Am J Physiol Cell Physiol*, 1996, 271, C1424–C1437.
8. Beishuizen ED, Tamsma JT, Jukema JW, van de Ree MA, van de Vijver JC, Meinders AE, Huisman MV: The effect of statin therapy on endothelial function in type 2 diabetes without manifest cardiovascular disease. *Diabetes Care*, 2005, 28, 1668–1674.
9. Beny JL, Schaad O: An evaluation of potassium ions as endothelium-derived hyperpolarizing factor in porcine coronary arteries. *Br J Pharmacol*, 2000, 131, 965–973.
10. Bjelakovic G, Nikolova D, Gluud LL, Simonetti RG, Gluud C: Antioxidant supplements for prevention of mortality in healthy participants and patients with various diseases. *Cochrane Database Syst Rev*, 2008, CD007176.
11. Boger RH: L-Arginine therapy in cardiovascular pathologies: beneficial or dangerous? *Curr Opin Clin Nutr Metab Care*, 2008, 11, 55–61.
12. Boo YC, Jo H: Flow-dependent regulation of endothelial nitric oxide synthase: role of protein kinases. *Am J Physiol Cell Physiol*, 2003, 285, C499–C508.
13. Burnham MP, Bychkov R, Félétou M, Richards GR, Vanhoutte PM, Weston AH, Edwards G: Characterization of an apamin-sensitive small-conductance Ca²⁺-activated K⁺ channel in porcine coronary artery endothelium: relevance to EDHF. *Br J Pharmacol*, 2002, 135, 1133–1143.
14. Buus NH, Simonsen U, Pilegaard HK, Mulvany MJ: Nitric oxide, prostanoid and non-NO, non-prostanoid involvement in acetylcholine relaxation of isolated human small arteries. *Br J Pharmacol*, 2000, 129, 184–192.
15. Cai H, Harrison DG: Endothelial dysfunction in cardiovascular diseases: the role of oxidant stress. *Circ Res*, 2000, 87, 840–844.
16. Carter TD, Ogden D: Acetylcholine-stimulated changes of membrane potential and intracellular Ca²⁺ concentration recorded in endothelial cells in situ in the isolated rat aorta. *Pflugers Arch*, 1994, 428, 476–484.
17. Celermajer DS: Endothelial dysfunction: does it matter? Is it reversible? *J Am Coll Cardiol*, 1997, 30, 325–333.

18. Ceriello A, Assaloni R, Da Ros R, Maier A, Piconi L, Quagliaro L, Esposito K et al.: Effect of atorvastatin and irbesartan, alone and in combination, on postprandial endothelial dysfunction, oxidative stress, and inflammation in type 2 diabetic patients. *Circulation*, 2005, 111, 2518–2524.
19. Ceriello A, Taboga C, Tonutti L, Quagliaro L, Piconi L, Bais B, Da Ros R et al.: Evidence for an independent and cumulative effect of postprandial hypertriglyceridemia and hyperglycemia on endothelial dysfunction and oxidative stress generation: effects of short- and long-term simvastatin treatment. *Circulation*, 2002, 106, 1211–1218.
20. Chen G, Cheung DW: Effect of K⁺-channel blockers on ACh-induced hyperpolarization and relaxation in mesenteric arteries. *Am J Physiol Heart Circ Physiol*, 1997, 272, H2306–H2312.
21. Chen Y, Pearlman A, Luo Z, Wilcox CS: Hydrogen peroxide mediates a transient vasorelaxation with tempol during oxidative stress. *Am J Physiol Heart Circ Physiol*, 2007, 293, H2085–H2092.
22. Chen YF, Cowley AW, Jr., Zou AP: Increased H₂O₂ counteracts the vasodilator and natriuretic effects of superoxide dismutation by tempol in renal medulla. *Am J Physiol Regul Integr Comp Physiol*, 2003, 285, R827–R833.
23. Chłopicki S, Gryglewski RJ: Angiotensin converting enzyme (ACE) and HydroxyMethylGlutaryl-CoA (HMG-CoA) reductase inhibitors in the forefront of pharmacology of endothelium. *Pharmacol Rep*, 2005, 57, Suppl, 86–96.
24. Christensen FH, Hansen T, Stankevicius E, Buus NH, Simonsen U: Elevated pressure selectively blunts flow-evoked vasodilatation in rat mesenteric small arteries. *Br J Pharmacol*, 2007, 150, 80–87.
25. Christensen FH, Stankevicius E, Hansen T, Jørgensen MM, Valverde VL, Simonsen U, Buus NH: Flow- and acetylcholine-induced dilatation in small arteries from rats with renovascular hypertension – effect of tempol treatment. *Eur J Pharmacol*, 2007, 566, 160–166.
26. Corson MA, James NL, Latta SE, Nerem RM, Berk BC, Harrison DG: Phosphorylation of endothelial nitric oxide synthase in response to fluid shear stress. *Circ Res*, 1996, 79, 984–991.
27. Didion SP, Kinzenbaw DA, Schrader LI, Faraci FM: Heterozygous CuZn superoxide dismutase deficiency produces a vascular phenotype with aging. *Hypertension*, 2006, 48, 1072–1079.
28. Dimmeler S, Fleming I, Fisslthaler B, Hermann C, Busse R, Zeiher AM: Activation of nitric oxide synthase in endothelial cells by Akt-dependent phosphorylation. *Nature*, 1999, 399, 601–605.
29. Dinkova-Kostova AT, Liby KT, Stephenson KK, Holtzclaw WD, Gao X, Suh N, Williams C et al.: Extremely potent triterpenoid inducers of the phase 2 response: correlations of protection against oxidant and inflammatory stress. *Proc Natl Acad Sci USA*, 2005, 102, 4584–4589.
30. Economides PA, Caselli A, Tian E, Khaodhiar L, Horton ES, Veves A: The effects of atorvastatin on endothelial function in diabetic patients and subjects at risk for type 2 diabetes. *J Clin Endocrinol Metab*, 2004, 89, 740–747.
31. Edwards G, Dora KA, Gardener MJ, Garland CJ, Weston AH: K⁺ is an endothelium-derived hyperpolarizing factor in rat arteries. *Nature*, 1998, 396, 269–272.
32. Estruch R, Martínez-González MA, Corella D, Salas-Salvadó J, Ruiz-Gutiérrez V, Covas MI, Fiol M et al.: Effects of a Mediterranean-style diet on cardiovascular risk factors: a randomized trial. *Ann Intern Med*, 2006, 145, 1–11.
33. Feng J, Liu Y, Clements RT, Sodha NR, Khabbaz KR, Senthilnathan V, Nishimura KK et al.: Calcium-activated potassium channels contribute to human coronary microvascular dysfunction after cardioplegic arrest. *Circulation*, 2008, 118, S46–S51.
34. Fleming I, Busse R: Molecular mechanisms involved in the regulation of the endothelial nitric oxide synthase. *Am J Physiol Regul Integr Comp Physiol*, 2003, 284, R1–R12.
35. Frobert O, Holmager P, Jensen KM, Schmidt EB, Simonsen U: Effect of acute changes in oxygen tension on flow-mediated dilation. Relation to cardiovascular risk. *Scand Cardiovasc J*, 2008, 42, 38–47.
36. Fukui T, Ishizaka N, Rajagopalan S, Laursen JB, Capers Q, Taylor WR, Harrison DG et al.: p22phox mRNA expression and NADPH oxidase activity are increased in aortas from hypertensive rats. *Circ Res*, 1997, 80, 45–51.
37. Fulton D, Gratton JP, McCabe TJ, Fontana J, Fujio Y, Walsh K, Franke TF et al.: Regulation of endothelium-derived nitric oxide production by the protein kinase Akt. *Nature*, 1999, 399, 597–601.
38. Gao X, Belmadani S, Picchi A, Xu X, Potter BJ, Tewari-Singh N, Capobianco S et al.: Tumor necrosis factor- α induces endothelial dysfunction in Lepr(db) mice. *Circulation*, 2007, 115, 245–254.
39. Garcia-Redondo AB, Briones AM, Beltran AE, Alonso MJ, Simonsen U, Salaices M: Hypertension increases contractile responses to hydrogen peroxide in resistance arteries through increased thromboxane A₂, Ca²⁺, and superoxide anion levels. *J Pharmacol Exp Ther*, 2009, 328, 19–27.
40. Ghiadoni L, Magagna A, Versari D, Kardasz I, Huang Y, Taddei S, Salvetti A: Different effect of antihypertensive drugs on conduit artery endothelial function. *Hypertension*, 2003, 41, 1281–1286.
41. Ghisda P, Morel N: Cellular target of voltage and calcium-dependent K⁺ channel blockers involved in EDHF-mediated responses in rat superior mesenteric artery. *Br J Pharmacol*, 2001, 134, 1021–1028.
42. Griendling KK, FitzGerald GA: Oxidative stress and cardiovascular injury: Part I: basic mechanisms and in vivo monitoring of ROS. *Circulation*, 2003, 108, 1912–1916.
43. Halcox JP, Schenke WH, Zalos G, Mincemoyer R, Prasad A, Waclawiw MA, Nour KR et al.: Prognostic value of coronary vascular endothelial dysfunction. *Circulation*, 2002, 106, 653–658.
44. Hamilton SJ, Chew GT, Watts GF: Therapeutic regulation of endothelial dysfunction in type 2 diabetes mellitus. *Diab Vasc Dis Res*, 2007, 4, 89–102.
45. Hernanz R, Alonso MJ, Zibrandtsen H, Alvarez Y, Salaices M, Simonsen U: Measurements of nitric oxide

- concentration and hyporeactivity in rat superior mesenteric artery exposed to endotoxin. *Cardiovasc Res*, 2004, 62, 202–211.
46. Hilgers RH, Webb RC: Reduced expression of SKCa and IKCa channel proteins in rat small mesenteric arteries during angiotensin II-induced hypertension. *Am J Physiol Heart Circ Physiol*, 2007, 292, H2275–H2284.
 47. Hwa JJ, Ghibaudi L, Williams P, Chatterjee M: Comparison of acetylcholine-dependent relaxation in large and small arteries of rat mesenteric vascular bed. *Am J Physiol Heart Circ Physiol*, 1994, 266, H952–H958.
 48. Iglarz M, Clozel M: Mechanisms of ET-1-induced endothelial dysfunction. *J Cardiovasc Pharmacol*, 2007, 50, 621–628.
 49. Klatt P, Heinzel B, Mayer B, Ambach E, Werner-Felmayer G, Wachter H, Werner ER: Stimulation of human nitric oxide synthase by tetrahydrobiopterin and selective binding of the cofactor. *FEBS Lett*, 1992, 305, 160–162.
 50. Köhler R, Degenhardt C, Kuhn M, Runkel N, Paul M, Hoyer J: Expression and function of endothelial Ca^{2+} -activated K^{+} channels in human mesenteric artery: A single-cell reverse transcriptase-polymerase chain reaction and electrophysiological study in situ. *Circ Res*, 2000, 87, 496–503.
 51. Kris-Etherton PM, Lichtenstein AH, Howard BV, Steinberg D, Witztum JL: Antioxidant vitamin supplements and cardiovascular disease. *Circulation*, 2004, 110, 637–641.
 52. Marchenko SM, Sage SO: Calcium-activated potassium channels in the endothelium of intact rat aorta. *J Physiol*, 1996, 492, 53–60.
 53. Marchesi C, Paradis P, Schiffrin EL: Role of the renin-angiotensin system in vascular inflammation. *Trends Pharmacol Sci*, 2008, 29, 367–374.
 54. Márquez Martín A, de la Puerta Vázquez R, Fernández-Arche A, Ruiz-Gutiérrez V: Suppressive effect of maslinic acid from pomace olive oil on oxidative stress and cytokine production in stimulated murine macrophages. *Free Radic Res*, 2006, 40, 295–302.
 55. McCabe TJ, Fulton D, Roman LJ, Sessa WC: Enhanced electron flux and reduced calmodulin dissociation may explain “calcium-independent” eNOS activation by phosphorylation. *J Biol Chem*, 2000, 275, 6123–6128.
 56. McSherry IN, Spitaler MM, Takano H, Dora KA: Endothelial cell Ca^{2+} increases are independent of membrane potential in pressurized rat mesenteric arteries. *Cell Calcium*, 2005, 38, 23–33.
 57. Mitchell JB, Samuni A, Krishna MC, DeGraff WG, Ahn MS, Samuni U, Russo A: Biologically active metal-independent superoxide dismutase mimics. *Biochemistry*, 1990, 29, 2802–2807.
 58. Modena MG, Bonetti L, Coppi F, Bursi F, Rossi R: Prognostic role of reversible endothelial dysfunction in hypertensive postmenopausal women. *J Am Coll Cardiol*, 2002, 40, 505–510.
 59. Moncada S: Nitric oxide in the vasculature: physiology and pathophysiology. *Ann N Y Acad Sci*, 1997, 811, 60–67.
 60. Montagnani M, Chen H, Barr VA, Quon MJ: Insulin-stimulated activation of eNOS is independent of Ca^{2+} but requires phosphorylation by Akt at Ser(1179). *J Biol Chem*, 2001, 276, 30392–30398.
 61. Munzel T, Sinning C, Post F, Warnholtz A, Schulz E: Pathophysiology, diagnosis and prognostic implications of endothelial dysfunction. *Ann Med*, 2008, 40, 180–196.
 62. Nilius B, Droogmans G: Ion channels and their functional role in vascular endothelium. *Physiol Rev*, 2001, 81, 1415–1459.
 63. Nitenberg A, Pham I, Antony I, Valensi P, Attali JR, Chemla D: Cardiovascular outcome of patients with abnormal coronary vasomotion and normal coronary arteriography is worse in type 2 diabetes mellitus than in arterial hypertension: a 10 year follow-up study. *Atherosclerosis*, 2005, 183, 113–120.
 64. Nitenberg A, Valensi P, Sachs R, Cosson E, Attali JR, Antony I: Prognostic value of epicardial coronary artery constriction to the cold pressor test in type 2 diabetic patients with angiographically normal coronary arteries and no other major coronary risk factors. *Diabetes Care*, 2004, 27, 208–215.
 65. Ozawa Y, Hayashi K, Kanda T, Homma K, Takamatsu I, Tatematsu S, Yoshioka K et al.: Impaired nitric oxide- and endothelium-derived hyperpolarizing factor-dependent dilation of renal afferent arteriole in Dahl salt-sensitive rats. *Nephrology (Carlton)*, 2004, 9, 272–277.
 66. Papapetropoulos A, Fulton D, Lin MI, Fontana J, McCabe TJ, Zoellner S, Garcia-Cardena G et al.: Vanadate is a potent activator of endothelial nitric-oxide synthase: evidence for the role of the serine/threonine kinase Akt and the 90-kDa heat shock protein. *Mol Pharmacol*, 2004, 65, 407–415.
 67. Parker RA, Huang Q, Tesfamariam B: Influence of 3-hydroxy-3-methylglutaryl-CoA (HMG-CoA) reductase inhibitors on endothelial nitric oxide synthase and the formation of oxidants in the vasculature. *Atherosclerosis*, 2003, 169, 19–29.
 68. Perez-Vizcaino F, Duarte J, Andriantsitohaina R: Endothelial function and cardiovascular disease: effects of quercetin and wine polyphenols. *Free Radic Res*, 2006, 40, 1054–1065.
 69. Perona JS, Arcemis C, Ruiz-Gutierrez V, Catala A: Effect of dietary high-oleic-acid oils that are rich in antioxidants on microsomal lipid peroxidation in rats. *J Agric Food Chem*, 2005, 53, 730–735.
 70. Perona JS, Cabello-Moruno R, Ruiz-Gutierrez V: The role of virgin olive oil components in the modulation of endothelial function. *J Nutr Biochem*, 2006, 17, 429–445.
 71. Perticone F, Ceravolo R, Maio R, Ventura G, Iacopino S, Cuda G, Mastroroberto P et al.: Calcium antagonist isradipine improves abnormal endothelium-dependent vasodilation in never treated hypertensive patients. *Cardiovasc Res*, 1999, 41, 299–306.
 72. Perticone F, Ceravolo R, Pujia A, Ventura G, Iacopino S, Scozzafava A, Ferraro A et al.: Prognostic significance of endothelial dysfunction in hypertensive patients. *Circulation*, 2001, 104, 191–196.
 73. Rodriguez-Rodriguez R, Herrera MD, de Sotomayor MA, Ruiz-Gutierrez V: Pomace olive oil improves endothelial function in spontaneously hypertensive rats by

- increasing endothelial nitric oxide synthase expression. *Am J Hypertens*, 2007, 20, 728–734.
74. Rodriguez-Rodriguez R, Herrera MD, Perona JS, Ruiz-Gutierrez V: Potential vasorelaxant effects of oleanolic acid and erythrodiol, two triterpenoids contained in 'orujo' olive oil, on rat aorta. *Br J Nutr*, 2004, 92, 635–642.
75. Rodriguez-Rodriguez R, Perona JS, Herrera MD, Ruiz-Gutierrez V: Triterpenic compounds from "orujo" olive oil elicit vasorelaxation in aorta from spontaneously hypertensive rats. *J Agric Food Chem*, 2006, 54, 2096–2102.
76. Rodriguez-Rodriguez R, Stankevicius E, Herrera MD, Ostergaard L, Andersen MR, Ruiz-Gutierrez V, Simonsen U: Oleanolic acid induces relaxation and calcium-independent release of endothelium-derived nitric oxide. *Br J Pharmacol*, 2008, 155, 535–546.
77. Russell KS, Haynes MP, Caulin-Glaser T, Rosneck J, Sessa WC, Bender JR: Estrogen stimulates heat shock protein 90 binding to endothelial nitric oxide synthase in human vascular endothelial cells. Effects on calcium sensitivity and NO release. *J Biol Chem*, 2000, 275, 5026–5030.
78. Samuni AM, DeGraff W, Krishna MC, Mitchell JB: Cellular sites of H₂O₂-induced damage and their protection by nitroxides. *Biochim Biophys Acta*, 2001, 1525, 70–76.
79. Sankaranarayanan A, Raman G, Busch C, Schultz T, Zimin PI, Hoyer J, Köhler R et al.: Naphtho[1,2-d]thiazol-2-ylamine (SKA-31), a new activator of KCa₂ and KCa_{3.1} potassium channels, potentiates the endothelium-derived hyperpolarizing factor response and lowers blood pressure. *Mol Pharmacol*, 2009, 75, 281–295.
80. Schachinger V, Britten MB, Zeiher AM: Prognostic impact of coronary vasodilator dysfunction on adverse long-term outcome of coronary heart disease. *Circulation*, 2000, 101, 1899–1906.
81. Schiffrin EL, Park JB, Intengan HD, Touyz RM: Correction of arterial structure and endothelial dysfunction in human essential hypertension by the angiotensin receptor antagonist losartan. *Circulation*, 2000, 101, 1653–1659.
82. Sheng JZ, Braun AP: Small- and intermediate-conductance Ca²⁺-activated K⁺ channels directly control agonist-evoked nitric oxide synthesis in human vascular endothelial cells. *Am J Physiol Cell Physiol*, 2007, 293, C458–C467.
83. Sheng JZ, Ella S, Davis MJ, Hill MA, Braun AP: Openers of SKCa and IKCa channels enhance agonist-evoked endothelial nitric oxide synthesis and arteriolar vasodilation. *FASEB J*, 2008, in press.
84. Shimokawa H, Yasutake H, Fujii K, Owada MK, Nakaike R, Fukumoto Y, Takayanagi T et al.: The importance of the hyperpolarizing mechanism increases as the vessel size decreases in endothelium-dependent relaxations in rat mesenteric circulation. *J Cardiovasc Pharmacol*, 1996, 28, 703–711.
85. Si H, Heyken WT, Wölfle SE, Tysiac M, Schubert R, Grgic I, Vilianovich L et al.: Impaired endothelium-derived hyperpolarizing factor-mediated dilations and increased blood pressure in mice deficient of the intermediate-conductance Ca²⁺-activated K⁺ channel. *Circ Res*, 2006, 99, 537–544.
86. Simonsen U, Wadsworth RM, Buus NH, Mulvany MJ: In vitro simultaneous measurements of relaxation and nitric oxide concentration in rat superior mesenteric artery. *J Physiol*, 1999, 516, 271–282.
87. Sommer F, Schulze W: Treating erectile dysfunction by endothelial rehabilitation with phosphodiesterase 5 inhibitors. *World J Urol*, 2005, 23, 385–392.
88. Somova LO, Nadar A, Rammanan P, Shode FO: Cardiovascular, antihyperlipidemic and antioxidant effects of oleanolic and ursolic acids in experimental hypertension. *Phytomedicine*, 2003, 10, 115–121.
89. Song M, Hang TJ, Wang Y, Jiang L, Wu XL, Zhang Z, Shen J et al.: Determination of oleanolic acid in human plasma and study of its pharmacokinetics in Chinese healthy male volunteers by HPLC tandem mass spectrometry. *J Pharm Biomed Anal*, 2006, 40, 190–196.
90. Stankevicius E, Hughes AD, Simonsen U: K channels and release of nitric oxide. *Fundam Clin Pharmacol*, 2008, s2, 22, 9.
91. Stankevicius E, Lopez-Valverde V, Rivera L, Hughes AD, Mulvany MJ, Simonsen U: Combination of Ca²⁺-activated K⁺ channel blockers inhibits acetylcholine-evoked nitric oxide release in rat superior mesenteric artery. *Br J Pharmacol*, 2006, 149, 560–572.
92. Stankevicius E, Martinez AC, Mulvany MJ, Simonsen U: Blunted acetylcholine relaxation and nitric oxide release in arteries from renal hypertensive rats. *J Hypertens*, 2002, 20, 1571–1579.
93. Strøbaek D, Teuber L, Jørgensen TD, Ahring PK, Kjaer K, Hansen RS, Olesen SP et al.: Activation of human IK and SK Ca²⁺-activated K⁺ channels by NS309 (6,7-dichloro-1H-indole-2,3-dione 3-oxime). *Biochim Biophys Acta*, 2004, 1665, 1–5.
94. Suwaidi JA, Hamasaki S, Higano ST, Nishimura RA, Holmes DR, Jr., Lerman A: Long-term follow-up of patients with mild coronary artery disease and endothelial dysfunction. *Circulation*, 2000, 101, 948–954.
95. Taddei S, Virdis A, Ghiadoni L, Magagna A, Favilla S, Pompella A, Salvetti A: Restoration of nitric oxide availability after calcium antagonist treatment in essential hypertension. *Hypertension*, 2001, 37, 943–948.
96. Tan KC, Chow WS, Tam SC, Ai VH, Lam CH, Lam KS: Atorvastatin lowers C-reactive protein and improves endothelium-dependent vasodilation in type 2 diabetes mellitus. *J Clin Endocrinol Metab*, 2002, 87, 563–568.
97. Taylor MS, Bonev AD, Gross TP, Eckman DM, Brayden JE, Bond CT, Adelman JP et al.: Altered expression of small-conductance Ca²⁺-activated K⁺ (SK3) channels modulates arterial tone and blood pressure. *Circ Res*, 2003, 93, 124–131.
98. Thorsgaard M, Lopez V, Buus NH, Simonsen U: Different modulation by Ca²⁺-activated K⁺ channel blockers and herbimycin of acetylcholine- and flow-evoked vasodilation in rat mesenteric small arteries. *Br J Pharmacol*, 2003, 138, 1562–1570.
99. Trichopoulou A, Costacou T, Bamia C, Trichopoulos D: Adherence to a Mediterranean diet and survival in a Greek population. *N Engl J Med*, 2003, 348, 2599–2608.

100. Tsunekawa T, Hayashi T, Kano H, Sumi D, Matsui-Hirai H, Thakur NK, Egashira K et al.: Cerivastatin, a hydroxymethylglutaryl coenzyme a reductase inhibitor, improves endothelial function in elderly diabetic patients within 3 days. *Circulation*, 2001, 104, 376–379.
101. Ungvari Z, Csiszar A, Huang A, Kaminski PM, Wolin MS, Koller A: High pressure induces superoxide production in isolated arteries via protein kinase C-dependent activation of NAD(P)H oxidase. *Circulation*, 2003, 108, 1253–1258.
102. Usachev YM, Marchenko SM, Sage SO: Cytosolic calcium concentration in resting and stimulated endothelium of excised intact rat aorta. *J Physiol*, 1995, 489, 309–317.
103. van Venrooij FV, van de Ree MA, Bots ML, Stolk RP, Huisman MV, Banga JD: Aggressive lipid lowering does not improve endothelial function in type 2 diabetes: the Diabetes Atorvastatin Lipid Intervention (DALI) Study: a randomized, double-blind, placebo-controlled trial. *Diabetes Care*, 2002, 25, 1211–1216.
104. Vásquez-Vivar J, Kalyanaraman B, Martásek P, Hogg N, Masters BS, Karoui H, Tordo P et al.: Superoxide generation by endothelial nitric oxide synthase: the influence of cofactors. *Proc Natl Acad Sci USA*, 1998, 95, 9220–9225.
105. Wulff H, Zhorov BS: K⁺ channel modulators for the treatment of neurological disorders and autoimmune diseases. *Chem Rev*, 2008, 108, 1744–1773.
106. Xiang L, Dearman J, Abram SR, Carter C, Hester RL: Insulin resistance and impaired functional vasodilation in obese Zucker rats. *Am J Physiol Heart Circ Physiol*, 2008, 294, H1658–H1666.
107. Xu H, Jackson WF, Fink GD, Galligan JJ: Activation of potassium channels by tempol in arterial smooth muscle cells from normotensive and deoxycorticosterone acetate-salt hypertensive rats. *Hypertension*, 2006, 48, 1080–1087.
108. Yada T, Shimokawa H, Morikawa K, Takaki A, Shinozaki Y, Mori H, Goto M et al.: Role of Cu,Zn-SOD in the synthesis of endogenous vasodilator hydrogen peroxide during reactive hyperemia in mouse mesenteric microcirculation in vivo. *Am J Physiol Heart Circ Physiol*, 2008, 294, H441–H448.
109. Yamamoto Y, Imaeda K, Suzuki H: Endothelium-dependent hyperpolarization and intercellular electrical coupling in guinea-pig mesenteric arterioles. *J Physiol*, 1999, 514, 505–513.
110. Zalba G, Beaumont FJ, San Jose G, Fortuno A, Fortuno MA, Etayo JC, Diez J: Vascular NADH/NADPH oxidase is involved in enhanced superoxide production in spontaneously hypertensive rats. *Hypertension*, 2000, 35, 1055–1061.
111. Zheng JS, Yang XQ, Lookingland KJ, Fink GD, Hesslinger C, Kapatos G, Kovsesdi I et al.: Gene transfer of human guanosine 5'-triphosphate cyclohydrolase I restores vascular tetrahydrobiopterin level and endothelial function in low renin hypertension. *Circulation*, 2003, 108, 1238–1245.
112. Zygmunt PM, Hogestatt ED: Role of potassium channels in endothelium-dependent relaxation resistant to nitroarginine in the rat hepatic artery. *Br J Pharmacol*, 1996, 117, 1600–1606.

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